

A PIPELINE FOR DNA-BASED IDENTIFICATION OF EXOTIC AND NATIVE MOSQUITOES IN BELGIUM

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2nd ANNUAL CONFERENCE

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■ BopCo: A Barcoding Facility for Organisms and Tissues of Policy Concern

- Policy concern organisms = endangered species, invasive alien species, human and veterinary disease organisms or their vectors, species of forensic relevance, agricultural pest species, organisms of the human food chain, etc.
- For all stakeholders, using 'traditional' morphological characteristics and/or DNA-based identification technology



MEMO: Monitoring of Exotic mosquito species



- Coordinated by the Institute of Tropical Medicine (Belgium);
- Nationwide monitoring project;
- Funded by the Belgian federal authorities;
- 3 years: July 2017- June 2020;
- Aim: detecting and evaluating the occurrence of exotic mosquito species (EMS), as they represent an increasing threat to human and animal health in temperate regions (main target: *Aedes*).

I. BACKGROUND



MEMO: Monitoring of Exotic mosquito species



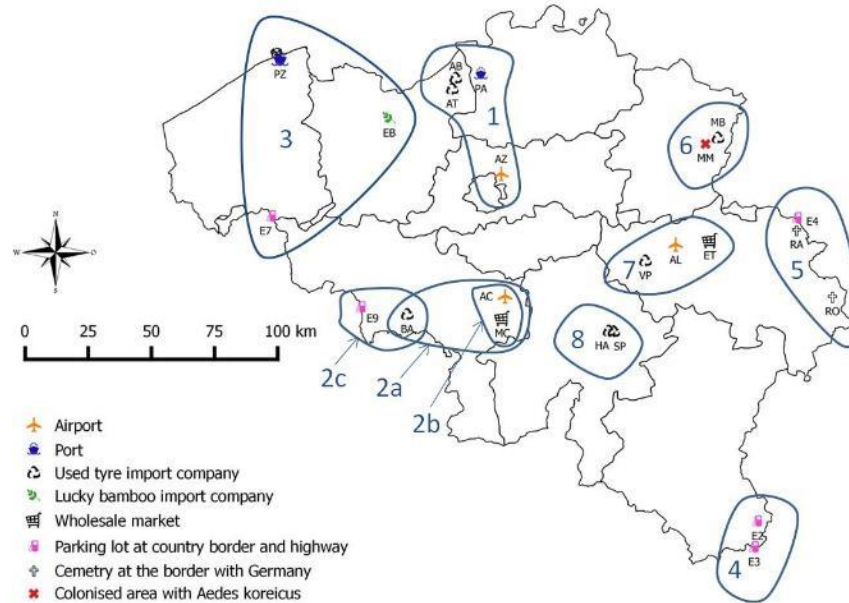
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→ **Critical step = Correct species identification**

I. BACKGROUND



Field work: 23 PoEs



-Eggs, larvae and adults collected using 14 Mosquito Magnet Independence Traps, 320 Oviposition Traps, 9 BG-Sentinel Traps.
 -146 potential breeding sites are investigated for larvae.



Known introduction pathways: lucky bamboo and used tyre import companies, port and airport, wholesale markets, natural spread.



I. BACKGROUND

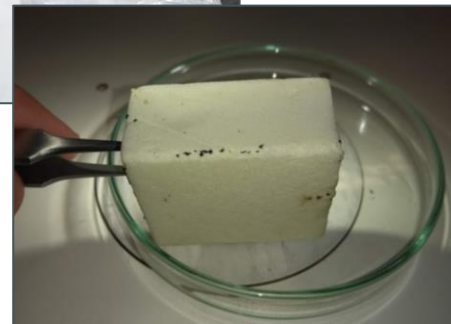


Morphological species identification



Specimens morphologically identified using:

- Digital identification key of Schaffner *et al.* (2001);
- Printed keys of Schaffner (1993) and Becker *et al.* (2010).



**INSTITUTE
OF TROPICAL
MEDICINE
ANTWERP**

I. BACKGROUND



Voucher collection

→ A few well preserved representatives per species selected: larvae on slides, adults pinned.



slide mounting of larvae



Royal Belgian
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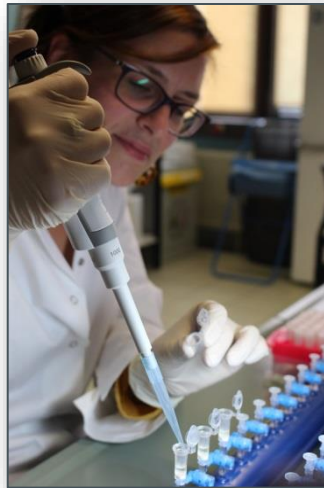


I. BACKGROUND



DNA-based species identification

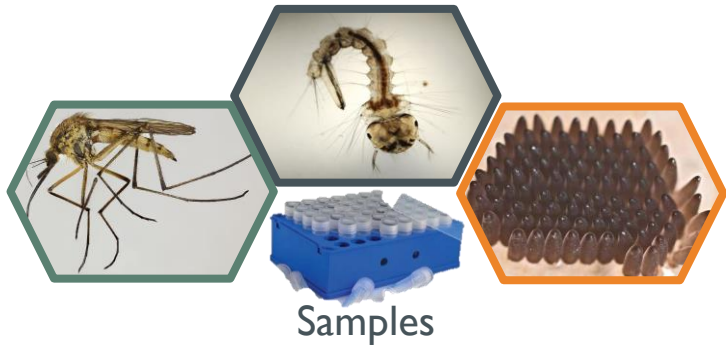
- **Ad hoc validation** of all (suspected) EMS and all specimens lacking discriminative characteristics (ex. early life stages, damaged or sp complex), mainly focusing on (native) *Aedes*;
- **Annual validation:** random selection of 5% of all caught specimens (adults and larvae) as quality control of the morphological species identifications.



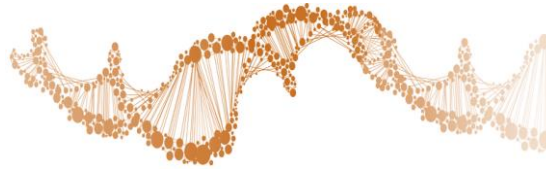
Gentegra technology for tissue and DNA storage



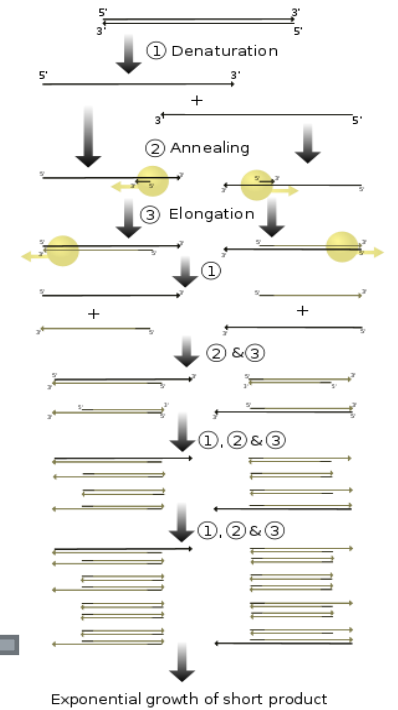
I. BACKGROUND: DNA BARCODING



DNA extraction and quantification

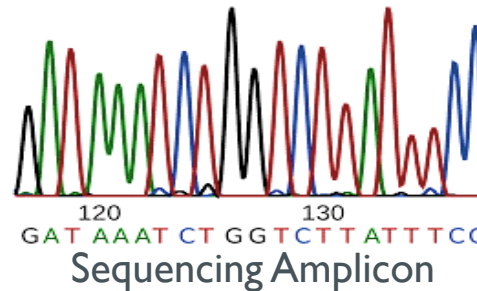
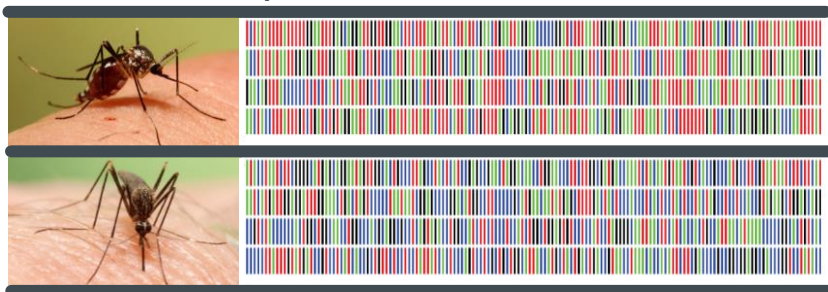


Amplification of selected fragments or loci (e.g. COI, NADH, ITS2, etc.)

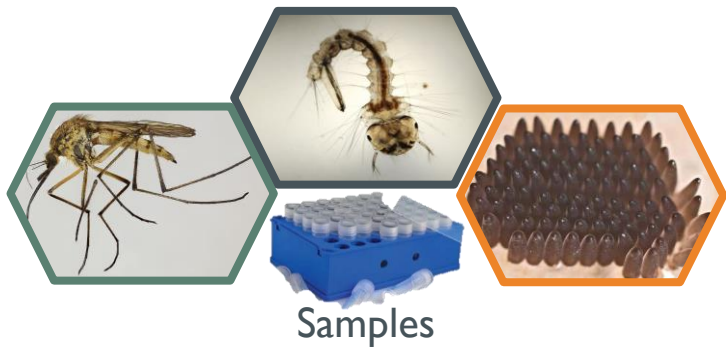


✓ Species identification

Online repositories database search



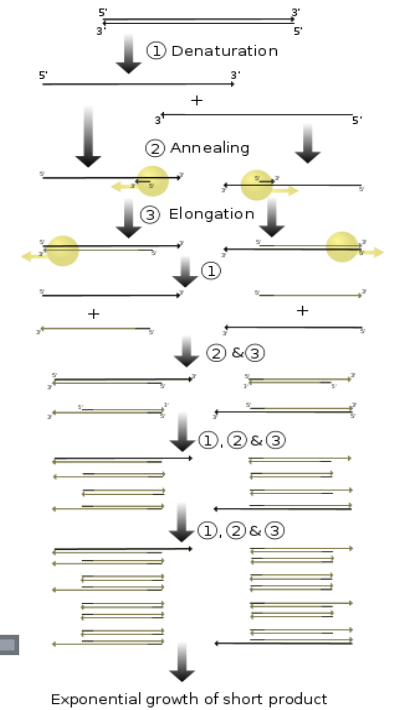
I. BACKGROUND: DNA BARCODING



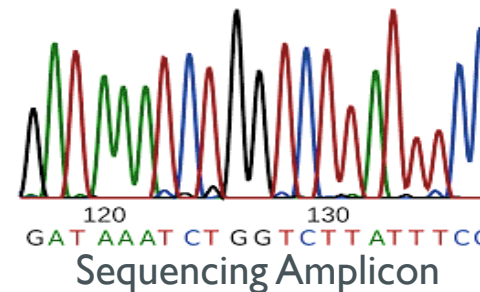
DNA extraction and quantification



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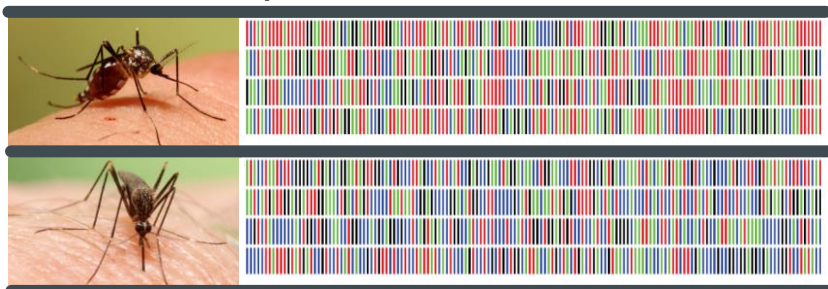


??? COMPLETENESS AND RELIABILITY ???



✓ Species identification

Online repositories database search



I. BACKGROUND: DNA BARCODING



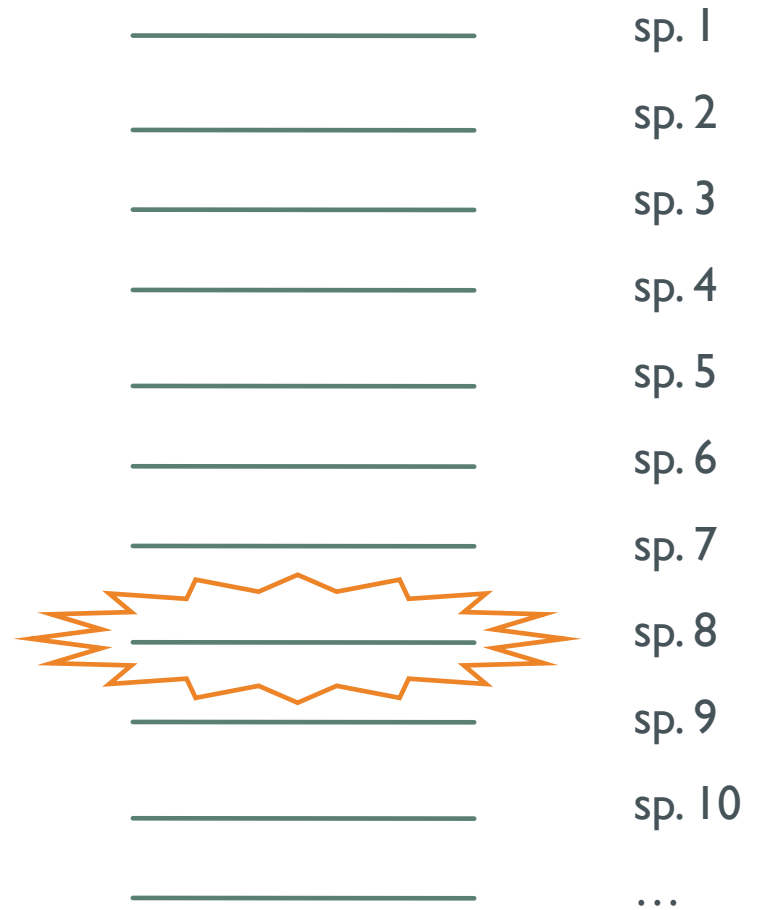
An ideal scenario:

inter-specific
>
intra-specific divergence

COMPLETE reference library
of DNA barcodes

unknown query
(sp. 8)

correct match!



Online DNA repository

I. BACKGROUND: DNA BARCODING



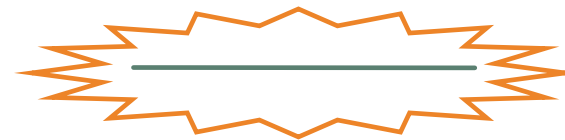
A more realistic scenario:

inter-specific
not necessarily >
intra-specific divergence

INCOMPLETE reference library
of DNA barcodes

unknown query
(sp. 8)

wrong match!
misidentification



- _____ sp. 1
 - _____ sp. 2 missing
 - _____ sp. 3
 - _____ sp. 4
 - _____ sp. 5 missing
 - _____ sp. 6 missing
 - _____ sp. 7
 - _____ sp. 8 missing
 - _____ sp. 9 missing
 - _____ sp. 10
 - _____ ...
- Online DNA repository

2. METHODS: **MOLECULAR SPECIES IDENTIFICATION WORKFLOW**



→ EVALUATE THE AVAILABLE REFERENCE LIBRARY OF DNA BARCODES
(BOLD & GenBank)

Genus	Species	Belgium →2020
Anopheles	<i>daciae</i>	✓
	<i>atroparvus</i>	✓
	<i>claviger</i>	✓
	<i>maculipennis</i>	✓
	<i>messeae</i>	✓
	<i>pharoensis</i>	✓
	<i>plumbeus</i>	✓
Aedes	<i>aegypti</i>	
	<i>triseriatus</i>	
	<i>cinereus</i>	✓
	<i>vexans</i>	✓
	<i>geniculatus</i>	✓
	<i>atropalpus</i>	
	<i>japonicus</i>	✓
	<i>koreicus</i>	✓
	<i>annulipes</i>	✓
	<i>cantans</i>	✓
	<i>caspius</i>	✓
	<i>communis</i>	✓
	<i>detritus</i>	✓
	<i>dorsalis</i>	✓
	<i>flavescens</i>	✓
	<i>punctor</i>	✓
	<i>riparius</i>	
	<i>sticticus</i>	✓
<i>rusticus</i>	✓	
<i>albopictus</i>	✓	

MOLECULAR SPECIES IDENTIFICATION WORKFLOW



➔ EVALUATE THE AVAILABLE REFERENCE LIBRARY OF DNA BARCODES (BOLD & GenBank)

- **Mosquito checklist:** literature review of culicid fauna of Belgium + **7 EMS of main concern to Central Europe** + updates from new records for Belgium (**3 native** and **1 EMS**);

Genus	Species	Belgium →2020
Culex	<i>pipiens</i>	✓
	<i>torrentium</i>	✓
	<i>quinquefasciatus</i>	
	<i>modestus</i>	✓
	<i>territans</i>	✓
	<i>hortensis</i>	✓
Culiseta	<i>fumipennis</i>	✓
	<i>morsitans</i>	✓
	<i>longiareolata</i>	✓
	<i>annulata</i>	✓
Coquillettidia	<i>subochrea</i>	✓
	<i>richiardii</i>	✓

**5 genera; 31 native species;
8 EMS of which 4 recorded in
Belgium before 2020**

2. METHODS: MOLECULAR SPECIES IDENTIFICATION WORKFLOW



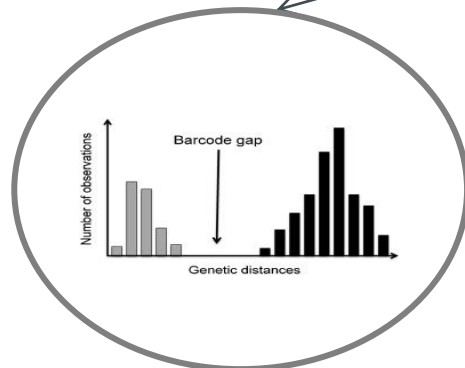
Download all sequence data available

Filtering the data

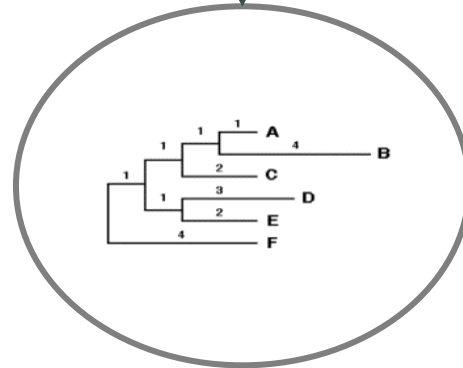
Aligning and trimming of the sequences

Database

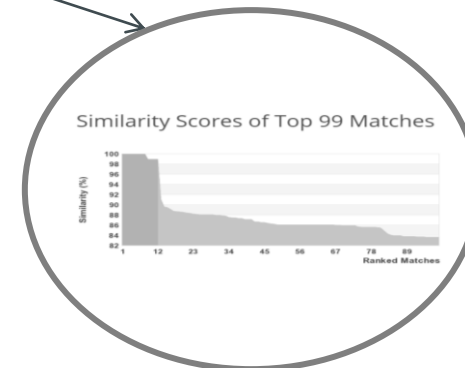
Include metadata like geographical origin of the sequenced voucher/specimen



Genetic distance (K2P-distance, p-distance)



Clustering using NJ-tree with Bootstrap support



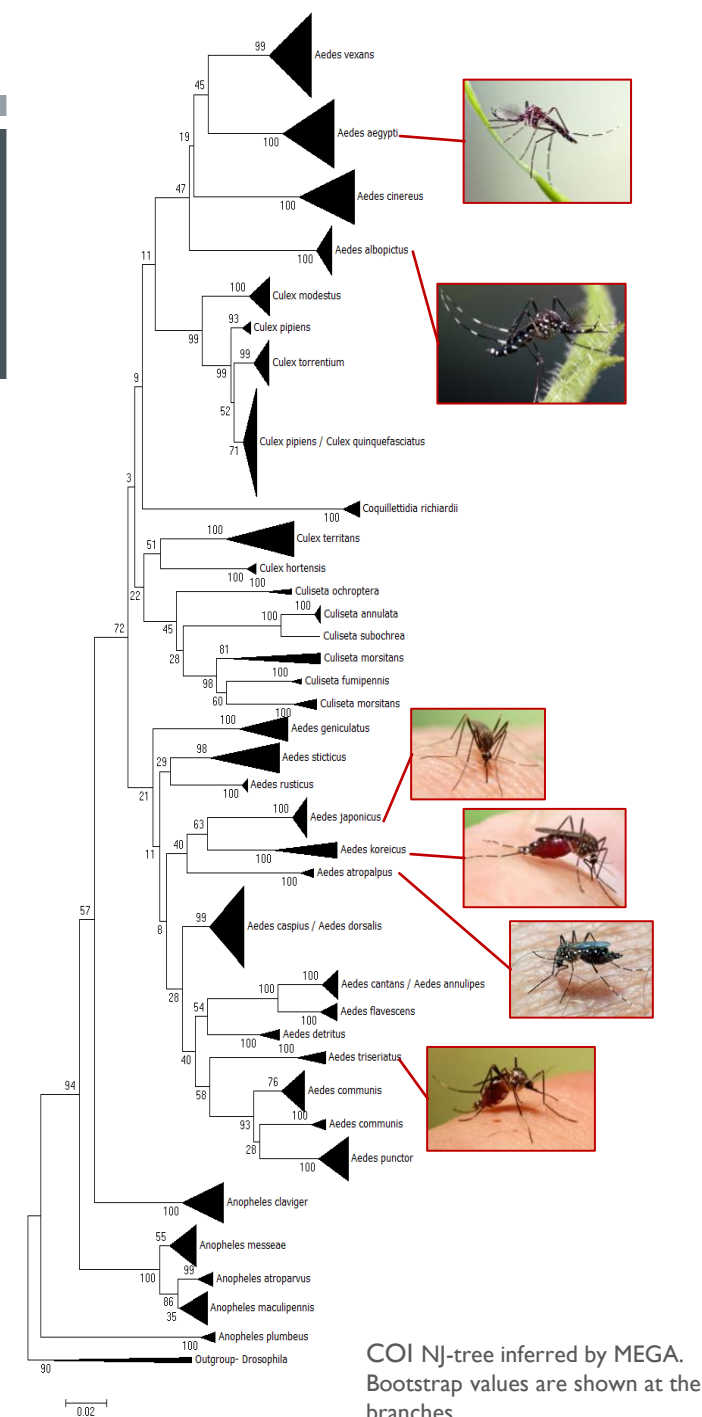
BLAST-IDS: Basic Local Alignment Tool (BLAST) in GenBank / BOLD

2. METHODS:

NJ-trees and barcoding gap analyses:

→ 26 of 39 culicid species identifiable using COI exclusively (including 7 EMS of the checklist);

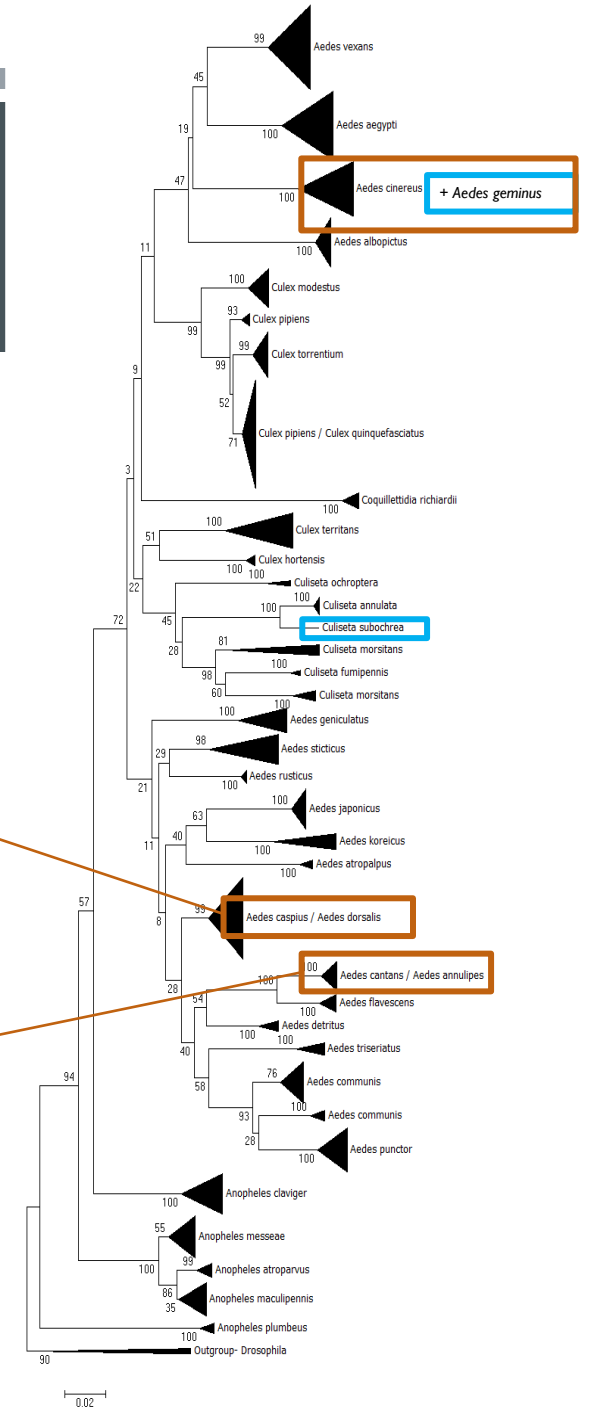
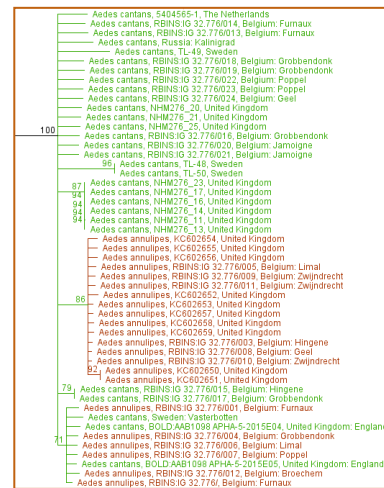
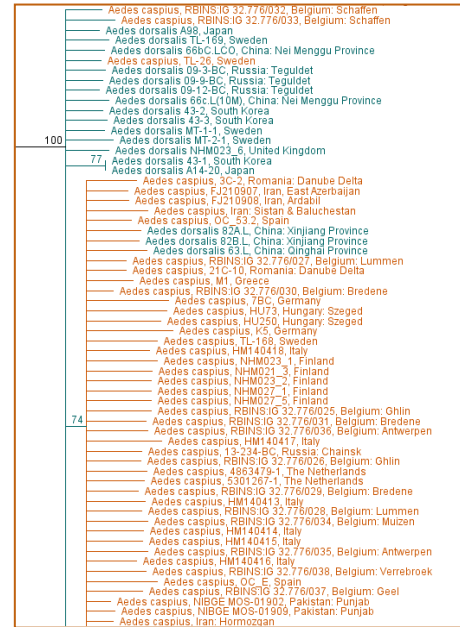
→ 8 other species identifiable using other DNA markers (ITS2, ACE2, CQ11 - including 1 EMS of the checklist);



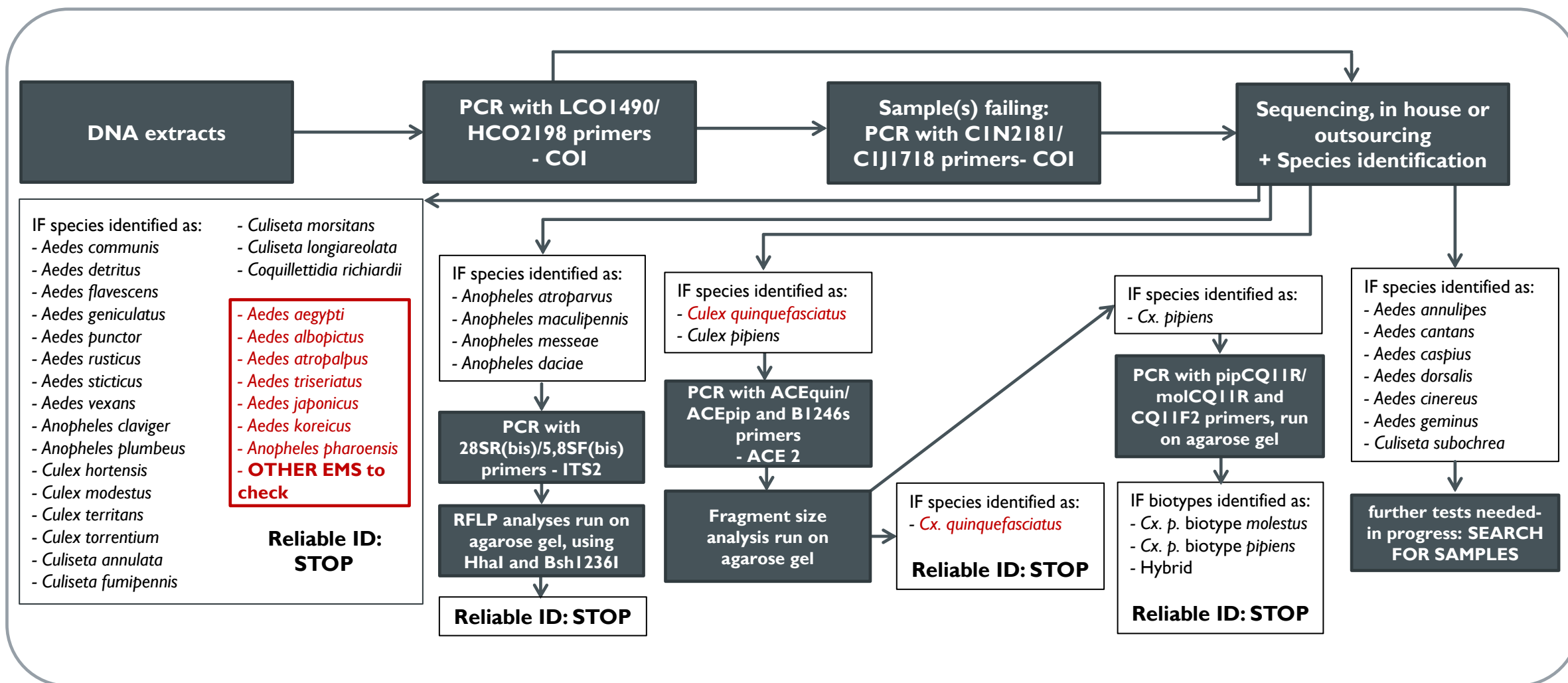
2. METHODS:

→ 3 species complexes
(*Aedes caspius* / *Ae. dorsalis*,
Ae. cantans / *Ae. annulipes*,
Ae. cinereus / *Ae. geminus*)

→ 2 species lacking
DNA data (*Aedes geminus*
& *Culiseta subochrea*).



2. METHODS: MOLECULAR SPECIES IDENTIFICATION WORKFLOW

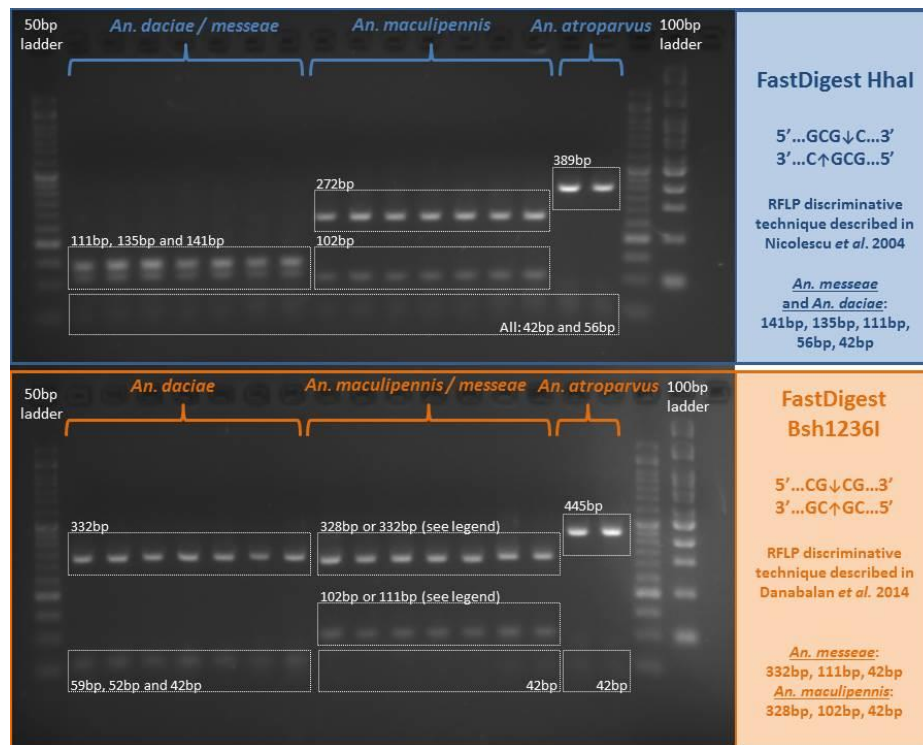


2. METHODS: MOLECULAR SPECIES IDENTIFICATION WORKFLOW

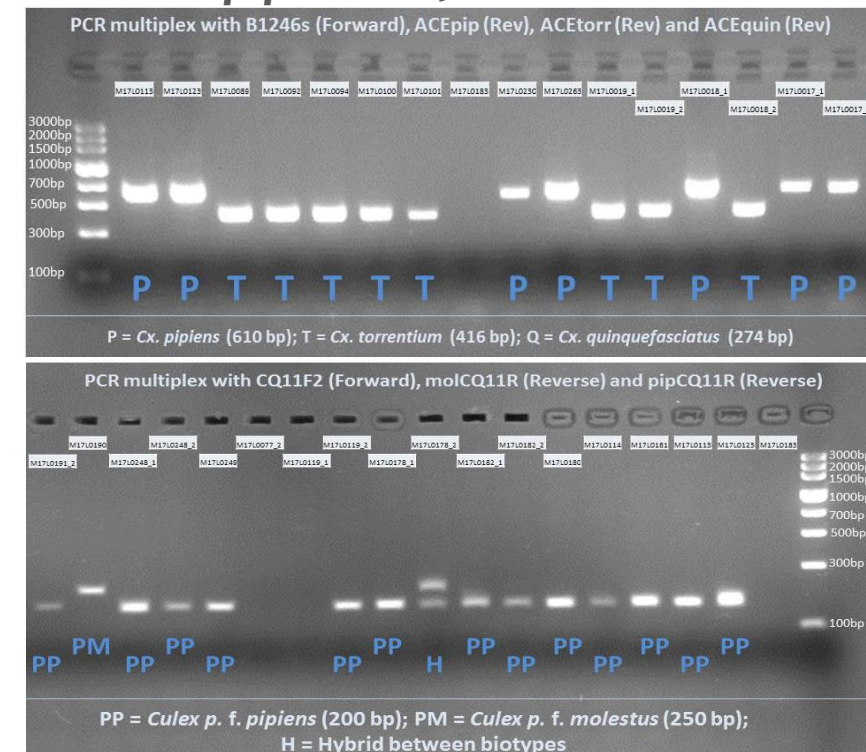


Size fragment analyses

→ *Anopheles maculipennis* s.l.



→ *Culex pipiens* s.l., *Cx. torrentium* and biotypes



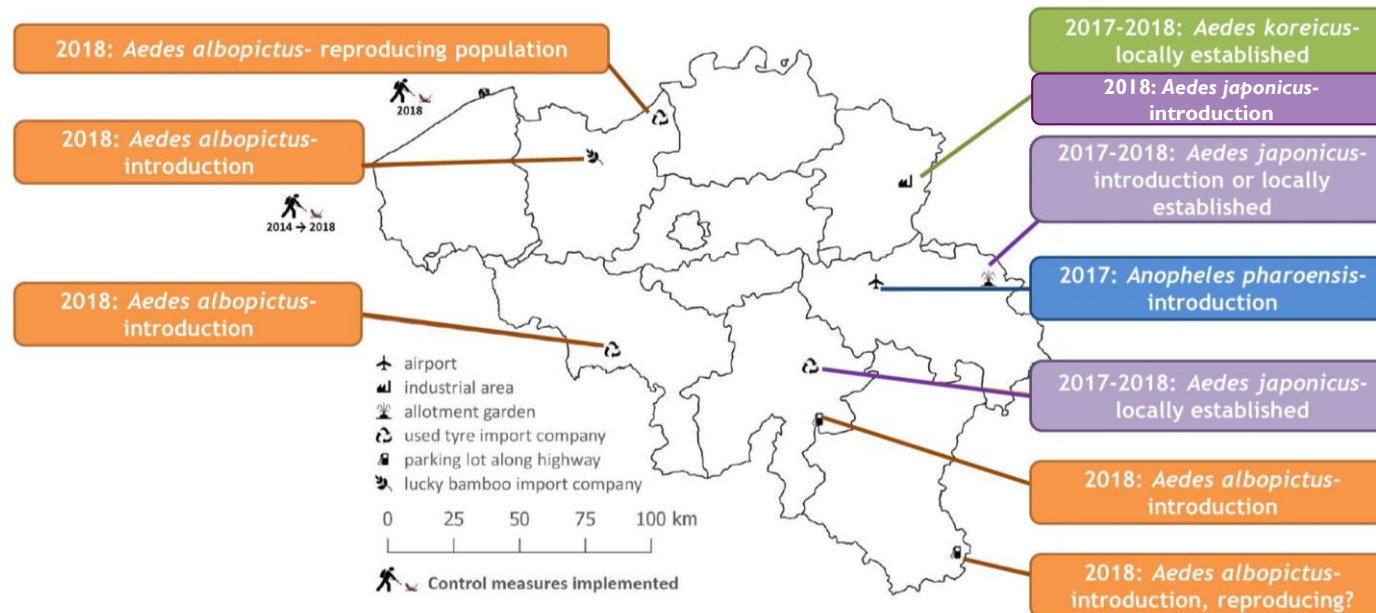
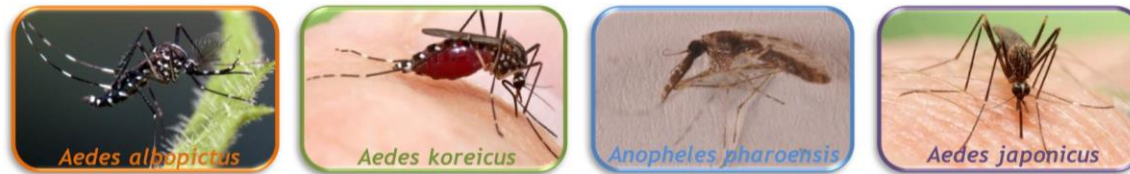
Alternative qPCR techniques optimized and tested (on progress)

3. RESULTS



Exotic mosquito species occurrence

Four EMS collected once or multiple times at one or multiple PoEs



→ Investigate introduction pathway through questionnaires.

→ Investigate possible intensification of the sampling in the surrounding area: established?

→ Investigate possible population genetic study(ies): source?

3. RESULTS



Ex. Annual validation (year 2018)

N = 1,285

5 genera

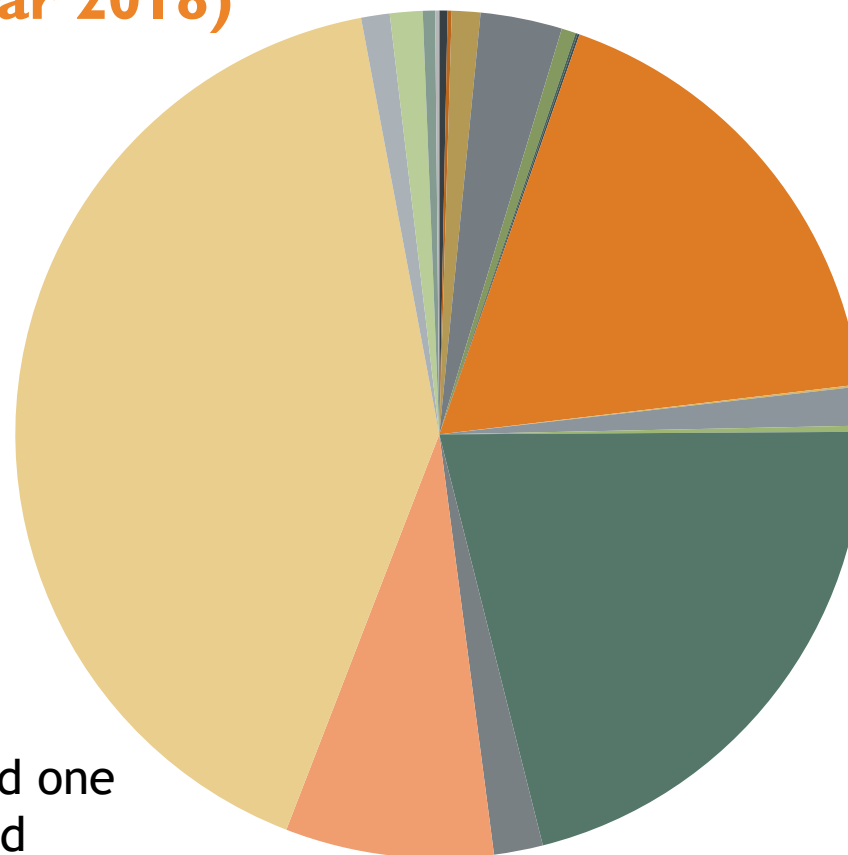
→ Most frequent species:

- *Culex pipiens*
- *Anopheles plumbeus*
- *Culex torrentium*

→ Most frequent biotype:

- *Cx. p.* form *pipiens*

→ Eleven **native**, two **exotic** and one **newly** recorded species validated



- *Aedes vexans* (N = 4)
- *Aedes cinereus* (N = 2)
- *Aedes geniculatus* (N = 14)
- *Aedes koreicus* (N = 40)
- *Aedes japonicus* (N = 7)
- *Anopheles maculipennis* (N = 1)
- *Anopheles daciae* (N = 1)
- *Anopheles plumbeus* (N = 228)
- *Anopheles claviger* (N = 1)
- *Culiseta annulata* (N = 19)
- *Culex hortensis* (N = 3)
- *Culex torrentium* (N = 272)
- *Culex pipiens s.l.* (N = 24)
- *Culex pipiens f. molestus* (N = 102)
- *Culex pipiens f. pipiens* (N = 529)
- *Culex pipiens s.s. (undetermined biotype)* (N = 14)
- *Hybrid Culex p. f. pipiens / Culex p. f. molestus* (N = 16)
- *Coquillettidia richiardii* (N = 6)
- No Identification (N = 2)

4. DISCUSSION



■ Annual validation:

- DNA-based identifications allow to validate the morphology-based species identifications (limits for 3 native sp complexes). Yet, it also allows to discriminate between cryptic species, or species of the same complex (*Cx. pipiens s.l.* / *Cx. torrentium*; *Anopheles maculipennis s.l.*).

Every year, more than 98% of the DNA-based identifications matched the respective morphological identifications,

- DNA-based techniques allow to validate/discover three new species records for Belgium: *Culiseta longiareolata*, *Culex modestus* and *Anopheles daciae*.

■ Ad hoc validation:

- 4 EMS collected at 9 distinct PoEs (2017-2018)
- Very useful in case of eggs (mainly with *Ae. albopictus* and *Ae. geniculatus*).

→ EMS seem to enter effectively via different introduction pathways.
Early interceptions and rapid DNA-based verifications should help the authorities in their efforts to contain the spread and eradicate EMS (populations).

QUESTIONS?



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A Barcoding Facility for Organisms and Tissues of Policy Concern

Search... SEARCH

All Taxonomy

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Welcome to BopCo

The **Barcoding Facility for Organisms and Tissues of Policy Concern** (BopCo) aims at developing an expertise forum to facilitate the identification of biological samples of policy concern. Such identifications can rely on traditional morphology-based approaches requiring taxonomic expertise and/or DNA-based techniques demanding specific skills and access to a fully equipped molecular laboratory.

The intent of BopCo therefore is **(1)** to act as a focal point for identifying biological materials upon request, using both morphological and DNA-based techniques, **(2)** to produce well-documented DNA barcodes of relevant taxa, **(3)** to maintain reference collections of barcoded organisms and the corresponding DNA barcode databases, and **(4)** to explore and implement new tools and techniques for species identification and DNA barcoding.

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